

Validation Report #029608

Summary

Antigen	Human Cystatin C (CST3)
Catalog number	ABIN365082
Supplier	Cusabio
Supplier catalog number	<u>CSB-E08384h</u>
Lot number	S29095913
Method validated	Enzyme-linked immunosorbent assay
Laboratory	Alamo Laboratories Inc
Validation number	<u>29608</u>
Positive Control	<u>Human urine</u>
Negative Control	Rat urine
Notes	Signal was detected in positive control sample and not in negative control sample.



Validation Date: 02/11/14

Full Methods

Primary Antibody

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Controls

- Positive control: Urine from normal adult human (specimen known to contain the target protein).
- Negative control: Urine from rat (specimens known to not contain the target protein).
- Standard curve: Serial two-fold dilutions from 500 ng/ml [500, 250, 125, 62.5, 31.2, 15.6, 7.8, 0] were generated from the standard provided in the kit using standard/sample diluent buffer.
- Spike control: Standard diluted in standard/PBS diluent buffer [62.5 and 0].

Protocol

- All reagents in the ELISA kit were brought up to room temperature (RT) before use.
- 100 µL of standard or sample were added to wells in ELISA plate pre-coated with capture antibody. All samples and standards were assayed in triplicate.
- The plate was covered with sealer (provided in kit) and incubated for 2 hours at 37°C. Unbound material was aspirated but the wells were NOT Washed.
- 100 µL of Biotin-Antibody (diluted 1:100 in "Biotin-Antibody Diluent") was added to each well. Plate was covered with sealer (provided in kit) and incubated for 1 hour at 37°C. Unbound Biotin-Antibody was removed from each well and plate was washed three times with 350 µL of wash buffer (provided in the kit). After the last wash the plate was inverted against clean absorbent paper to remove any remaining liquid.
- 100 μL of HRP-Avidin Conjugate (diluted 1:100 in "HRP-Avidin Diluent") was added to each well. Plate was covered with sealer (provided in kit) and incubated for 1 hour at 37°C.
- Unbound HRP-Avidin was removed by washing five times with 350 µL of wash buffer (provided in the kit). After the last wash the plate was inverted and blotted against clean absorbent paper to remove any remaining liquid.
- 90 μL of TMB substrate was added to wells and the plate was covered with a new plate sealer. The plate was gently tapped to ensure mixing and incubated for 30 min at 37°C in the dark.
- After 30 min, when an apparent gradient appeared in the standard wells, the reaction was terminated by adding 50 µL of Stop Solution to each well.
- The optical density (OD value) of each well was read using a microplate reader set to 450 nm.
- The triplicate readings for each sample were averaged and the average zero standard optical density subtracted to yield 'corrected absorbance at 450 nm'. A standard curve was generated by plotting the mean OD value for each standard on the X-axis against the concentration on the Y-axis using Excel. Standard curve was generated by regression analysis with four-parameter logistic.
- An equation (y = -1106.2x4 + 2631.5x3 1044.9x2 + 234.72x 1.4717) was derived from the standard curve and used to calculate CST3 concentrations in samples based on their Average Absorbance values.

Experimental Notes

None

Figures

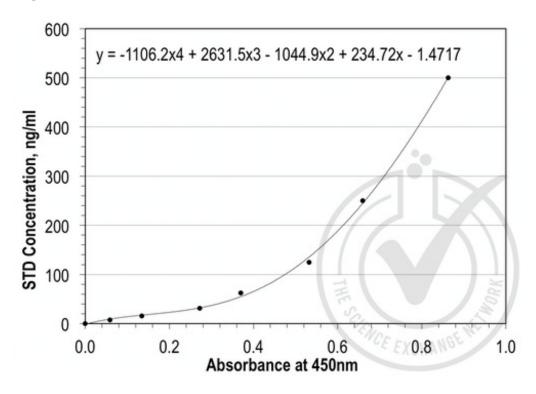


Figure 1: Graph of corrected OD 450 nm plotted for standard curve samples.

Туре	Sample, ng/ml	Readings at 450 nm			Avg Reading	Corrected OD450nm	SD	Calculated conc ng/ml
		1	2	3	Rea	9 9	0,	Calcu
Standards	500	0.998	0.946	0.993	0.979	0.863	0.023	500.65
	250	0.812	0.629	0.885	0.775	0.659	0.108	244.25
	125	0.584	0.531	0.829	0.648	0.532	0.130	135.28
	62.5	0.484	0.484	0.489	0.486	0.370	0.002	54.78
	31.2	0.353	0.412	0.400	0.388	0.272	0.025	32.02
	15.6	0.255	0.220	0.277	0.251	0.135	0.023	17.25
	7.8	0.152	0.187	0.185	0.175	0.059	0.016	9.22
	0	0.117	0.113	0.118	0.116	0.000	0.002	-1.47
Spike Controls	0.00	0.119	0.136	0.099	0.118	0.002	0.015	-1.01
	62.50	0.512	0.508	0.489	0.503	0.387	0.010	60.58
Test Samples	Urine Human	0.633	0.597	0.642	0.624	0.508	0.019	119.43
	Urine, Rat	0.145	0.138	0.143	0.142	0.026	0.003	3.97

Conc of Cystatin-C in Human Urine (+ ve Control): 119.43 ng/ml Conc of Cystatin-C in Rat Urine (-ve Control): 3.97 ng/ml

Table 1: ELISA. Cystatin C (CST3) is present in human urine and undetectable in rat urine. Spike controls indicate no interference in absorbance readings from the diluent used to prepare standards and sera samples. Absorbance readings (OD 450 nm) are shown for standard curve, spike controls and unknown samples. Value for Avg Reading is derived from the average reading of three samples. Avg Reading for "0" amount of Standard was subtracted from all Avg Readings to yield "Corrected OD450 nm" values for Standards, spike controls and unknown samples. Standard deviation is included for all samples. Standard curve was generated by regression analysis with four-parameter logistic. An equation (y = -1106.2x4 + 2631.5x3 - 1044.9x2 + 234.72x - 1.4717) was derived from the standard curve and used to calculate CST3 concentrations shown in Table 1.